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Convenient Synthesis of Pyruvate Acetals of Carbohydrates by Coupling of Trialkylsilylated Diols and Pyruvates

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Abstract: Hexopyranoside diols, mainly 4,6-diols with α -gluco, β -gluco, α -galacto, β -galacto, and α -manno configurations could be converted effectively (40 - 74% yields), to the corresponding pyruvate acetals by the coupling of the 0-trialkylsilylated, namely, 0-TBDMS and/or 0-TMS diols, and ethyl pyruvate in the presence of TMSOTf at temperatures between -30 °C and +3 °C. In the case of the β -manno isomer, the anomerization of the β -glycoside to the α -glycoside as well as the ring contraction of the pyranoside to the furanoside predominated.

INTRODUCTION

Pyruvic acid acetal is a universal acidic component of bacterial polysaccharides and frequently found in extracellular polysaccharides¹ of Gram-negative bacteria, *i.e.*, capsular polysaccharides, and occasionally in cell wall lipopolysaccharides¹ and in lipooligosaccharides.² Pyruvic acid acetal of hexopyranosides has proven to be very useful tool for immunochemical studies of Klebsiella polysaccharides.³ Acetalization of carbohydrate diol with pyruvate by direct condensation or acetal exchange in the presence of acid is supposed to have difficulty due to the adjacent electron-withdrawing carboxylate group and such unsuccessful attempts were indicated in the papers.^{4,5} Therefore, several indirect methods for the acetalization were reported. The first pyruvate acetal was prepared by catalytic oxidation of 4,6-acetal of 1-hydroxy-2-propanone.⁶ albeit in less than 10% yield. Recently ruthenium tetraoxide oxidation of furan-2-ylethylidene⁷ and 1-(3,4-dimethoxyphenyl)ethylidene⁸ intermediates which were obtained by transacetalization proved to give the desired pyruvate acetal in good yields. Further, transacetalization⁹ of methyl pyruvate diphenyl dithioacetal, catalyzed by thiophilic reagents, was also reported. A new approach to this problem, namely, the pyruvate formation from the silvlated carbohydrate diol and alkyl pyruvate in the presence of trimethylsilyl triflate (TMSOTf) has proven to be the method of choice by our ¹⁰ and other^{4,11} groups, and has been employed in the synthesis of oligosaccharides containing the pyruvic acid acetal.¹² In this paper the detail and scope of this acetalization using trialkylsilylated diols reported preliminary for some years ago¹⁰ are described.

RESULTS AND DISCUSSION

In order to study the scope of this acetalization reaction two different trialkylsilyl groups, namely, tertbutyldimethylsilyl and trimethylsilyl, were employed for the activation of hexopyranoside 4,6-diols. Six 2,3di-O-benzylated hexopyranosides having α -gluco (1a), α -galacto (2a), α -manno (3a), β -gluco (4a), β -galacto (5a), and β -manno (6a) configurations were used as 4,6-diols. Among these, cyclohexyl β -D-glucopyranoside (4a) and cyclohexyl β -D-mannopyranoside (6a) were each synthesized from the corresponding α -glycosides, 1 b and 3a, respectively, by a similar series of reactions: acetolysis, hydrobrominolysis and glycosylation with silver silicate as shown in the scheme (1b \rightarrow 7 \rightarrow 9 \rightarrow 11 \rightarrow 4a and 3a \rightarrow 8 \rightarrow 10 \rightarrow 12 \rightarrow 6a).

Treatment of the 4,6-diols of the gluco (1a and 4a) and manno (3a and 6a) configurations with tert-butylchlorodimethylsilane (TBDMSCI) and imidazole in N, N-dimethylformamide (DMF) at room temperature gave the corresponding 4,6-O-bis-TBDMS derivatives (1c, 3c, 4c and 6c) each in over 80% yield. However, the same silylation of the 4,6-diols (2a and 5a) of the galacto configuration afforded the 6-O-TBDMS derivatives (2d and 5d), which were characterized as the 4-acetates (2e and 5e) and further treated with chlorotrimethylsilane to provide the 6-O-TBDMS-4-O-TMS derivatives (2f and 5f) in yields of 90% and 95%, respectively.



O-Trimethylsilylation with chlorotrimethylsilane in the same manner was not successful presumably due to hydrolysis of the 6-O-TMS ether during the extraction with water to remove imidazole. The 4,6-bis-O-trimethylsilylhexopyranosides (1 g, 3 g, 4 g, and 6 g) could be obtained in yields of over 80% by treatment of the

diols (1a, 3a, 4a, and 6a) with 1,1,1,3,3,3-hexamethyldisilazane in the presence of trifluoroacetic acid in dichloromethane. All these bis-O-trialkylsilyl derivatives were sufficiently stable to survive chromatographic purification on a short column of silica gel using ether.

The acetalization reaction of pyruvate with trialkylsilylated diol in the presence of TMSOTf proved to be dependent on the type of silyl groups employed, the configuration of the hexopyranoside as well as the configuration of the anomeric position.



Acetalization using the TBDMS ethers proved to be an efficient process. Namely, the reaction of a disilylated hexopyranoside (1 mmol) with ethyl pyruvate (2 mmol) in the presence of TMSOTF (0.4 mmol) in dichloromethane or ether at -5 - +3 °C for one – several days provided the 4,6-acetal in about 50 – 70% yield except the case of the β -manno isomer 6 c (Table 1). Although the reaction proceeded more slowly in ether than in dichloromethane, the total yield of the two stereoisomers remained relatively unchanged in the two solvents. The high stereoselectivities were observed in the cases of α -manno (in dichloromethane) and β -galacto isomers, where the corresponding (S)-acetals were formed in 100% and 35% diastereomer excess, respectively.

Entry	Disilvl		Solvent	Temp.	Time	Product and Yield (%)				
,	ether		DUITUR	(°C)	(d)	4,6-Acetal	R	S	Total	
1	α-gluco	1c	CH ₂ Cl ₂	-5	2	13	31	24	55	
2	a-gluco	lc	Et ₂ O	-5	2	13	18	28	46	
3	a-galacto	2f	CH ₂ Cl ₂	-5	2	14	33	32	65	
4	a-galacto	2f	Et ₂ O	-5	2	14	36	27	63	
5	α-manno	3c	CH ₂ Cl ₂	-5	3	15	0	60 ^d	60	
6	α-manno	3c	Et ₂ O	-5	5	15	28	34 ^d	62	
7	β-gluco	4 c	CH ₂ Cl ₂	-5	4	17	32	26	58	
8	β-gluco	4c	Et ₂ O	+3	2	17	24	31	55	
9	β-galacto	5f	CH ₂ Cl ₂	-5	1	18	42	20	62	
10	β-galacto	5f	ELO	-5	2	18	35	19	54	
11	β-manno	6c	CH ₂ Cl ₂	-5	3	16	<11 °	11 ^d	<22	
1 2	β-manno	6c	Et ₂ O	-5	6	16	<10 °	11 ^d	<21	

Table 1. Pyruvate^a acetalation with TBDMS ethers^b in the presence of TMSOTf.^c

a: Ethyl pyruvate (2 equiv) was used. b: 4,6-Bis-O-TBDMS ethers were used in the cases of *gluco* and *manno* isomers, while 6-O-TBDMS-4-O-TMS ethers in the cases of *galacto* isomers. c: 40 Mol% (0.4 equiv) TMSOTf was used as catalyst. d: Estimated by ¹H-NMR signal and containing the 5,6-acetal **20** (3-6%). e: Contaminated with small amount of unidendified product.

These acetals having the axially oriented alkoxycarbonyl group were shown¹¹ to be the more thermodynamically stable based on the fact that the (R)-pyruvate acetal of the α -gluco isomer could be epimerized to the corresponding (S)-pyruvate acetal in the presence of TMSOTf at room temperature in dichloromethane. For all the reactions performed in ether the more thermodynamically stable isomers, *i.e.*, (S)-acetals for the gluco and manno¹³ configurations, (R)-acetals for the galacto configurations, were found to slightly predominate. The reactions of gluco isomers in dichloromethane displayed a reversed selectivity resulting in acetals possessing the equatorially oriented ethoxycarbonyl group predominantly.¹⁴

The configuration of the acetal carbon atom of the 4,6-acetals 13-19 was determined by ¹H- and ¹³C-NMR chemical shifts of the acetalic methyl groups (Table 2), by comparison with those previously reported.¹⁵

In the case of β -D-manno isomer 6c the desired 4,6-acetal 19 could not be obtained. Instead, its α -anomer 16 and the 5,6-acetal 20 were formed both in low yields. The structure of 20 was suggested by the ¹³C-NMR chemical shifts of the acetalic methyl (22.70 ppm) and quaternary carbon (105.59 ppm), which did not correspond to those of pyranosides. The furanoside structure was confirmed by the anomeric carbon chemical shift (104.23 ppm) and by the coupling constants of the ring protons measured at 500 MHz, indicating an equilibrium between the ²T₃ and ₂E conformers.

The formation of 20 can be explained by a competitive attack on the intermediary carbenium ion 22 by the ring oxygen (path b) as shown in the following scheme of the acetalization mechanism. The transition state resulting in the β -manno-4,6-acetal may be energetically elevated due to the axial substituent at C-2. In addition, the cleavage of C1-O ring bond may be accelerated due to the $\Delta 2$ effect.¹⁶ Therefore, acetalization using the less sterically hindered TMS ether, which should also be a more reactive diol derivative, attracted our interest.

Pyruvate		δ ¹ H(C-CH ₃)		δ ¹³ C(C- <i>C</i> H ₃)	δ ¹³ C(C-CH ₃)		
acetai		• R	S	R	S	R	S	
4,6-acetal								
a-Glc	13	1.67	1.55	17.88	25.46	97.68	98.98	
α-Glc ^a		1.71	1.56	17.5	25.3			
α-Gal	14	1.64	1.56	26.00	20.37	98.76	97.19	
α-Gal 🛚		1.59	1.66	25.8	18.3			
α-Man	15	1.72	1.52	17.88	25.62	98.00	99.30	
α-Man	16	1.72	1.57	17.82	25.57	97.89	99.25	
β-Glc	17	1.68	1.55	17.82	25.46	97.51	98.92	
β-Glc [*]		1.70	1.56	17.6	25.2			
β-Gal	18	1.65	1.58	25.90	23.35	98.76	96.97	
β-Gal *		1.60		25.7	-			
β-Man	19	1.70	-	17.93		97.95		
5,6-acetal								
α-Manf	20	1.57		22	.70	105.59		

Table 2. ¹H- and ¹³C-NMR chemical shifts of acetalic and adjacent methyl carbons in CDCl₃

a: Reported data for methyl 4,6-O-(1-methoxycarbonylethylidene)-D-hexopyranosides.¹⁵



First of all, the difference of reactivity between the TBDMS and TMS ethers was examined using the α gluco (1 g) and α -manno (3 g) isomers. The higher reactivity of these TMS ethers was displayed by the fact that the reaction proceeded effectively using less amount (0.2 equiv) of TMSOTf and at lower temperatures. While the 4,6- α -manno-acetal 15 was isolated in 46 - 64% yield together with the 5,6-acetal 21 (5 - 13%), the 4,6- α gluco-acetal 13 was obtained in about 40% yield (Table 3).¹⁷

			Solvent	TMSOTf	Temp.	Time		Product and Yield (%)					
Entry	Disilyl							4,6-Acetal					
	Clinci			((R	S	Total	Other	acetal	
1	α-gluco	1g	Et ₂ O	20	-30-→-20	3+6	13	18	22	40	29a	trace	
2	α-manno	3g	CH ₂ Cl ₂	40	-30	2	15	28	18 ^b	46	21	9 ^b	
3	α-manno	3g	CH ₂ Cl ₂	20	-20	2	15	25	21 ^b	46	21	13 ^b	
4	α-manno	3g	Et ₂ O	40	-20-→-5	4+1	15	29	24 ^b	53	21	10 ^b	
5	α-manno	3g	Et ₂ O	20	-5	2	15	31	33 ^b	64	21	5 ^b	
6	β-gluco	4g	CH ₂ Cl ₂	40	-30- →-20	1+1	17	12	12	24	20 °		
7	β-gluco	4g	Et ₂ O	40	-20-→-5	1+3	17	34	40	74	_		
8	β-manno	6g	CH ₂ Cl ₂	40	-20	2	16	< 9 ^{b,0}	ⁱ 10 ^b	< 19	20	17 ^b	
9	β-manno	6g	Et ₂ O	40	-20	4	19	5	0	17 °	20	21 ^b	

Table 3. Pyruvate^a acetalation with TMS ethers in the presence of TMSOTf

a: Ethyl pyruvate was (2 equiv) was used. b: Estimated by ¹H-NMR signal. c: A mixture of 6,6'acetal (10%) and its unidentified regioisomer (10%). d: Comtaminated with small amount of unidentified product. e: The α -isomer 16 was also obtained (12%).

This acetalization procedure was successfully applied to the β -gluco isomer 4 g, to give the 4,6-acetal 17 in 74% yield. In general, the desired 4,6-acetals were obtained in better yields in ether than in dichloromethane, indicating that the susceptibility of TMS ether to acid induces some side reactions described above and seems to be more easily controlled in ether. In contrast, the acetalization of 6 g gave the desired 4,6-acetal 19 in low yield, while the 5,6-acetal 20 was obtained as the major product.¹⁸

After our preliminary communication¹⁰ extensive studies on synthesis of 4,6-pyruvate acetals using silvlated hexopyranosides in the presence of TMSOTf were published.^{4,11} In comparison with these reports the most characteristic feature of our acetalization conditions seems to be the lower reaction temperature. The first effect of low reaction temperature appears to suppress the undesired anomerization and ring contraction of βglycopyranosides such as 23 and 24 to the furanosides, which were recently reported by Ziegler et al.¹¹ Under our conditions 4g and 5f did not give any rearranged products even in dichloromethane. On the contrary, the acetalization of 4 g at room temperature gave, after 7 h, the expected 4,6-acetal 17 in 29% yield [27% (S)-isomer and 2% (R)-isomer together with the α -anomer of 17S (9%) and the 5,6-acetal, *i.e.*, D-gluco isomer of 20 (6%). In the case of 5f, after 9 h the anomerized 4,6-acetal 14R was obtained predominantly in 10% yield together with very small amount of the desired 4,6-acetal 18 and the corresponding 5,6-acetal (10%). The second effect of the low reaction temperature is on the stereoselectivity of the 4,6-acetal formation. While at ambient temperature^{4,11} the thermodynamically favored isomer is obtained predominantly, at the lower temperature (-30°C--5°C) it is possible to get the kinetically favored isomers as the major product depending on reaction conditions. The third effect is an induction time, which was suggested by the observation that the reaction started all at once after a day or a few days and is complete within another day or two. Among the three steps involved in this acetalization (I~III in the scheme of acetal formation) the late-determining step is thought to be the second one (II), the formation of the carbenium ion. As previously reported glycosylidenation, the trimethylsilyloxy group is considered at first to be eliminated as 1,1,1,3,3,3-hexamethyldisiloxane by the attack of TMSOTf. It was found very recently that addition of small amount of trifluoromethanesulfonic acid (TfOH) accelerated the acetalization.19



In addition to the 4,6-acetal, the pyruvate acetals of vicinal diols have been found in nature. Although we reported the preparation of 3,4-pyruvate acetal (26) of α -D-galactopyranoside in the preliminary communication^{10,20}, the reported yield was incorrect and actually very low, giving also almost the same amount of an unexpected acetal²¹ 27. Namely, methyl 2,6-di-*O*-benzyl-3,4-bis-*O*-(trimethylsilyl)- α -D-galactopyranoside (25) was coupled with ethyl pyruvate in the presence of TMSOTf and TfOH in ether to give 26 and 27 in 13 % and 18 % yields, respectively. More efficient preparation of 26 could be carried out by a modified method using 1-acetoxy-2-propanone and will be reported elsewhere.

The characteristics of this pyruvate acetalization are summarized as follows. (1) Both TBDMS and TMS ethers of hexopyranoside 4,6-diols having α -gluco, β -gluco, α -galacto, β -galacto, and α -manno configurations can be acetalated to give the corresponding 4,6-acetals in about 50 – 70% yield. (2) In the cases of TMS ethers the acetalization is recommended to be carried out in ether in the presence of lower amount (20 mol%) of TMSOTf in order to prevent the side reactions. (3) The acetalization reaction in ether gives predominantly a pyruvate acetal having an equatorially oriented methyl group, that is, (S)-acetal for gluco and manno isomers and (R)-acetal for galacto one. However, pyruvate acetal having an axially oriented methyl group can be obtained as major product depending on the reaction conditions. (4) The acetalization of the β -manno isomer gave the 5,6-acetal predominantly. In conclusion, the pyruvate acetalization of trialkylsilylated carbohydrate diols in the presence of TMSOTf proved to be one of practical method, which is applicable widely to various types of naturally occurring pyruvic acid acetals.

EXPERIMENTAL

General methods. Melting points were measured with a Yanagimoto micro melting point apparatus and are uncorrected. Solutions were evaporated under diminished pressure at a bath temperature not exceeding 50 °C. Optical rotations were measured in a 0.5-dm tube with a JASCO DIP-4 polarimeter for solutions in chloroform, unless stated otherwise. IR spectra were recorded with a Hitachi model EPI-G2 spectrometer. ¹H- and ¹³C-NMR spectra were recorded with JEOL JNM-PS100, JNM-FX90Q, EX-270, and GX-500 spectrometers for solutions in chloroform-d containing tetramethylsilane as a internal reference.

Cyclohexyl 4,6-di-O-acetyl-2,3-di-O-benzyl- β -D-glucopyranoside (11). To a stirred solution of 1,4,6-tri-O-acetyl-2,3-di-O-benzyl- α -D-glucopyranose²² (7, 743 mg, 1.52 mmol) in dry dichloromethane (0.5 ml) was added a solution 25 % hydrogen bromide in acetic acid (3.3 ml) at 0 °C. The mixture was diluted with

chloroform after the disappearance of 7, washed twice with cold water, cold aqueous sodium hydrogencarbonate, and cold water again. The organic layer was dried over anhydrous sodium sulfate and concentrated to give 4,6-di-O-acetyl-2,3-di-O-benzyl- α -D-glucopyranosyl bromide (9, 770 mg), whose structure was confirmed by ¹H-NMR data: δ 7.40-7.21 (m, 10H, 2Ph), 6.32 (d, $J_{1,2}$ 4.0 Hz, H-1), 5.08 (t, $J_{3,4} = J_{4,5}$ 9.6 Hz, H-4), 4.89, 4.64 (ABq, J 12.0 Hz, PhCH₂), 4.70 (s, 2H, PhCH₂), 4.52-3.86 (m, 4H), 3.57 (dd, $J_{2,3}$ 9.8 Hz, H-2), 2.05, 1.93 (each s, each 3H, OAc).

To a solution of freshly distilled cyclohexanol (173 mg, 1.73 mmol) in dry toluene (2 ml) was added silver silicate (765 mg) with stirring at -15 °C. After 15 min, to the stirred mixture was added dropwise a solution of 9 prepared from 7 (743 mg, 1.52 mmol) in dry toluene (3.5 ml) at the same temperature for 12 min. After 30 min the mixture was brought to room temperature, diluted with chloroform and insoluble materials were filtered off. The organic layer was dried and evaporated, and the residue was purified by chromatography on silica gel with hexane-ethyl acetate (6:1) to give 11 (394 mg, 65 % from 7) as a white crystal; mp 94.0-95.0 °C; $[\alpha]_D$ -21.8° (c 1.1, CHCl₃); IR v_{max} (NaCl) 1735 cm⁻¹ (ester); ¹H-NMR δ 7.46-7.26 (m, 10H, 2Ph), 5.07 (t, $J_{3,4} = J_{4,5}$ 8.0 Hz, H-4), 4.90, 4.67 (ABq, J 12.0 Hz, PhCH₂), 4.90, 4.67 (ABq, J 10.0 Hz, PhCH₂), 4.64-4.56 (m, 1H, H-1), 4.33 (dd, $J_{5,6a}$ 5.0 Hz, J_{gem} 12.0 Hz, H-6a), 4.12 (dd, $J_{5,6b}$ 3.0 Hz, H-6b), 3.90-3.46 (m, 4H), 2.08, 1.92 (each s, each 3H, 2OAc), 2.18-1.14 (m, 10H, 5CH₂); ¹³C-NMR δ 170.76, 169.57 (C=O), 138.36, 128.34-127.64 (2Ph), 102.01 (d, J_{C-H} 158.7 Hz, C-1), 81.97, 81.80, 78.17, 75.09, 74.98, 71.73, 70.16, 62.73, 33.64, 31.96, 25.62, 23.95, 20.75. Anal. Calcd. for C₃₀H₃₈O₈: C, 68.42; H, 7.27. Found: C, 68.77; H, 7.25.

Cyclohexyl 4,6-di-O-acetyl-2,3-di-O-benzyl- β -D-mannopyranoside (12). Compound 12 was prepared from 1,4,6-tri-O-acetyl-2,3-di-O-benzyl- α -D-mannopyranose²³ (8) in the same manner as described for 11. Compound 8 prepared by acetolysis of methyl 2,3-di-O-benzyl- α -D-mannopyranoside²⁴ (3a) was converted to the corresponding α -bromide²⁵ 10, whose glycosylation using silver silicate gave the β -glucoside 12. The crude 12 was recrystallized from hexane-ethyl acetate to give white crystals (70 % from 8); mp 87.0-89.0 °C; [α]_D-81.9° (c 1.1, CHCl₃); IR ν _{max} (NaCl) 1728 cm⁻¹ (ester); ¹H-NMR δ 7.57-7.14 (m, 10 H, 2Ph), 5.33 (t, $J_{3,4} = J_{4,5}$ 8.0 Hz, H-4), 5.00, 4.84 (ABq, J 12.4 Hz, PhCH₂), 4.51 (s, 1H, H-1), 4.48, 4.29 (ABq, J 12.4 Hz, PhCH₂), 4.29-4.10 (m, 1H, H-6a), 3.90-3.34 (m, 5H), 2.05, 2.01 (each s, each 3H, 2OAc), 2.18-1.10 (m, 10H, 5CH₂). Anal. Calcd. for C₃₀H₃₈O₈: C, 68.42; H, 7.27. Found: C, 68.07; H, 7.37.

Cyclohexyl 2,3-di-*O*-benzyl-β-D-glucopyranoside (4a). To a solution of 11 (382 mg, 0.72 mmol) in 70 % aqueous methanol (20 ml) was added triethylamine (2.0 ml) with stirring. The mixture was allowed to stand overnight and concentrated. The residue was purified by chromatography on silica gel with hexane-ethyl acetate (1:1) to give 4a (296 mg, 93 %) as a white crystal; mp 112.0-113.0 °C; $[\alpha]_D$ -31.5° (c 1.0, CHCl₃); IR v_{max} (NaCl) 3550-2950 cm⁻¹ (OH); ¹H-NMR δ 7.39-7.23 (m, 10H, 2Ph), 4.97, 4.77 (ABq, J 11.0 Hz, PhCH₂), 4.92, 4.77 (ABq, J 11.0 Hz, PhCH₂), 4.92, 4.77 (ABq, J 11.0 Hz, PhCH₂), 4.60-4.45 (br, 1H, H-1), 3.91-3.17 (m, 7H), 2.98-2.31 (br, 2H, OH-4,6), 2.09-1.11 (m, 10H, 5CH₂); ¹³C-NMR δ 138.63, 138.47, 128.50-127.69 (2Ph), 101.96 (d, J_C H 158.7 Hz, C-1), 84.13, 81.86, 77.90, 75.14, 74.92, 74.71, 70.54, 62.63 (C-6), 33.80, 31.96, 25.57, 24.05, 23.95. Anal. Calcd. for C₂₆H₂₄O₆: C, 70.56; H 7.74. Found: C, 70.16; H, 7.34.

Cyclohexyl 2,3-di-O-benzyl-β-D-mannopyranoside (6a). Compound **6a** was prepared from 12 in the same manner as described for **4a**. The crude **6a** was recrystallized from ethyl acetate-ether to give white crystals (87%); mp 103.5-104.5 °C; $[\alpha]_D$ -125.0° (c 1.1, CHCl₃); IR v_{max} (NaCl) 3600-3100 cm⁻¹ (OH); ¹H-NMR δ 7.53-7.25 (m, 10H, 2Ph), 5.04, 4.85 (ABq, J 12.0 Hz, PhCH₂), 4.64 (d, J_{1,2} 1.8 Hz, H-1), 4.54, 4.32 (ABq, J 11.0 Hz, PhCH₂), 4.13-3.32 (m, 7H), 2.82-2.31 (br, 2H, OH-4,6), 2.08-1.07 (m, 10H, 5CH₂); ¹³C-NMR δ 138.52, 137.77, 128.45-127.53 (2Ph), 99.52 (d, J_{C-H} 156.3 Hz, C-1), 81.70, 77.25, 76.71, 75.90, 74.06, 71.13, 67.23, 62.73 (C-6), 33.43, 31.53, 25.62, 23.73, 23.62. Anal. Calcd. for C₂₆H₃₄O₆: C, 70.56; H, 7.74. Found: C, 70.69; H, 7.93.

Bis-O-tert-butyldimethylsilylation of 1a, 3a, 4a, and 6a. To a dried diols $(1a, 2^2 3a, 4a, or 6a, 1.0 mmol)$ was added a solution of *tert*-butylchlorodimethylsilane (3.0 mmol) and imidazole (3.1 mmol) in dry N,N-dimethylformamide (DMF, 2.3 ml) and allowed to stand for one day. The mixture was diluted with ether, washed with water, and dried over anhydrous sodium sulfate. The organic layer was concentrated and the residue was purified on a short column of silica gel with ether to give silylated products 1 c, 3 c, 4 c, and 6 c, respectively.

Methyl 2,3-di-O-benzyl-4,6-bis-O-(tert-butyldimethylsilyl)-α-D-glucopyranoside (1c): Yield quantitative; $[\alpha]_D$ -50.1° (c 2.0, CHCl₃); IR ν_{max} (NaCl) 1255 cm⁻¹ (SiMe); ¹H-NMR δ 7.32-7.12 (m, 10H, 2Ph), 5.02, 4.69 (ABq, J 11.0 Hz, PhCH₂), 4.67, 4.52 (ABq, J 11.5 Hz, PhCH₂), 4.64 (d, $J_{1,2}$ 3.0 Hz, H-1), 3.91-3.26 (m, 6H), 3.39 (s, 3H, OMe), 0.90 (s, 18H, 2CMe₃), 0.06, 0.04 (each s, each 6H, 2SiMe₂); ¹³C-NMR δ 139.21, 138.08, 128.34-127.01 (2Ph), 97.45 (C-1), 81.67, 80.74, 75.02, 73.12, 72.74, 70.75, 62.59, 54.75 (OMe), 25.95, 25.86 (2CMe₃), 18.35, 18.06 (2CMe₃), -3.83, -4.78, -5.07, -5.37 (2SiMe₂). Anal. Calcd. for C₃₃H₅₄O₆Si₂: C, 65.74; H, 9.03. Found: C, 65.59; H, 9.09.

Methyl 2,3-di-O-benzyl-4,6-bis-O- (*tert*-butyldimethylsilyl)- α -D-mannopyranoside (3c): Yield 80%; $[\alpha]_D$ -28.1° (c 1.4, CHCl₃); IR v_{max} (NaCl) 1253 cm⁻¹ (SiMe); ¹H-NMR δ 7.39-7.15 (m, 10H, 2Ph), 4.69 (d, $J_{1,2}$ 2.0 Hz, H-1), 4.58, 4.52 (each s, 4H, 2PhCH₂), 4.07-3.37 (m, 6H), 3.31 (s, 3H, OMe), 0.88, 0.86 (each s, each 9H, 2CMe₃), 0.07 (s, 12H, 2SiMe₂); ¹³C-NMR δ 138.62, 128.19-127.26 (2Ph), 98.87 (C-1), 80.50, 74.61, 72.63, 71.45, 67.94, 63.07, 54.43 (OMe), 25.98, 25.91 (2CMe₃), 18.38, 18.19 (2CMe₃), 3.90, -4.82, -5.07, -5.25 (2SiMe₂). Anal. Calcd. for C₃₃H₅₄O₆Si₂: C, 65.74; H, 9.03. Found: C, 65.37; H, 8.97.

Cyclohexyl 2,3-di-O-benzyl-4,6-bis-O-(tert-butyldimethylsilyl)-β-D-glucopyranoside (4c): Yield quantitative; $[\alpha]_D$ +23.5° (c 1.7, CHCl₃); IR v_{max} (NaCl) 1254 cm⁻¹ (SiMe); ¹H-NMR δ 7.34-7.14 (m, 10H, 2Ph), 4.99, 4.65 (ABq, J 11.8 Hz, PhCH₂), 4.97, 4.56 (ABq, J 10.4 Hz, PhCH₂), 4.74-4.56 (m, 1H, H-1), 3.99-3.11 (m, 7H), 2.12-1.12 (m, 10H, 5CH₂), 0.92, 0.86 (each s, each 9H, 2CMe₃), 0.10, 0.08, 0.04, 0.01 (each s, each 3H, 2SiMe₂); ¹³C-NMR δ 139.15, 138.66, 128.26-127.12 (2Ph), 101.88 (d, J_{C-H} 153.8 Hz, C-1), 84.81, 82.92, 77.39, 74.85, 74.52, 70.78, 62.77, 33.84, 31.99, 24.09, 25.93, 25.71 (2CMe₃), 18.40, 18.13 (2CMc₃), -3.81,-4.79 (2SiMe₂). Anal. Calcd. for C₃₈H₆₂O₆Si₂: C, 68.01; H, 9.31. Found: C, 68.05; H, 9.20.

Cyclohexyl 2,3-di-O-benzyl-4,6-bis-O- (tert-butyldimethylsilyl)-β-D-mannopyranoside (6c): Yield quantitative ; $[α]_D$ -50.6° (c 1.7, CHCl₃); IR v_{max} (NaCl) 1254 cm⁻¹ (SiMe); ¹H-NMR δ 7.48-7.14 (m, 10H, 2Ph), 4.95, 4.70 (ABq, J 11.6 Hz, PhCH₂), 4.50, 4.32 (ABq, J 11.2 Hz, PhCH₂), 4.50 (s, 1H, H-1), 4.02-3.08 (m, 7H), 2.00-1.18 (m, 10H, 5CH₂), 0.90, 0.86 (each s, each 9H, 2CMe₃), 0.06, 0.01 (each s, each 6H, 2SiMe₂); ¹³C-NMR δ 139.37, 128.09-126.99 (2Ph), 99.39 (C-1), 82.41, 78.44, 76.19, 74.32, 73.64, 70.66, 68.12, 63.07, 33.52, 25.77, 23.83, 18.17, 25.97, 25.82 (2CMe₃), 18.17 (2CMe₃) -3.79, -4.92, -5.23, -5.32 (2SiMe₂). Anal. Calcd. for C₃₈H₆₂O₆Si₂: C, 68.01; H, 9.31. Found: C, 67.93; H,9.25.

Bis-O-trimethylsilylation of 1a, 3a, 4a, and 6a. To a stirred solution of **1a, 3a, 4a, and 6a** (1.0 mmol) and 1,1,1,3,3,3-hexamethyldisilazane (2.0 mmol) in dry dichloromethane (3.2 ml) was added 2 drops of trifluoroacetic acid. After 4 - 23 h, the mixture was concentrated and the residue was purified on a column of silica gel with hexane-ethyl acetate (10:1 or 15:1) to give 1g, 3g, 4g, and 6g, respectively.

Methyl 2,3-di-O-benzyl-4,6-bis-O-trimethylsilyl- α -D-glucopyranoside (1g): Yield 84%; mp 42.5-45.0°C (petroleum ether); $[\alpha]_D$ +45.1° (c 1.7, CHCl₃); ¹H-NMR δ 7.28-7.23 (m, 10H, 2Ph), 5.00, 4.72 (ABq, J 11.2 Hz, PhCH₂), 4.70, 4.54 (ABq, J 12.0 Hz, PhCH₂), 4.58 (d, J_{1,2} 4.0 Hz, H-1), 3.90-3.30 (m, 6H), 3.36 (s, 3H, OMe), 0.10 (s, 18H, 2SiMe₃). Anal. Calcd. for C₂₇H₄₂O₆Si₂: C, 62.51; H, 8.16. Found: C, 62.45; H, 8.43.

Methyl 2,3-di-O-benzyl-4,6-bis-O-trimethylsilyl-α-D-mannopyranoside (3g): Yield 92%; $[α]_D$ +39.3° (c 1.4, CHCl₃); IR v_{max} (NaCl) 1245 cm⁻¹ (SiMe); ¹H-NMR δ 7.28-7.04 (m, 10H, 2Ph), 4.59 (d, $J_{1,2}$ 2.0 Hz, H-1), 4.58, 4.44 (ABq, J 12.0 Hz, PhCH₂), 4.45 (s, 2H, PhCH₂), 3.97-3.24 (m, 6H), 3.20 (s, 3H, OMe), 0.04, 0.01 (each s, each 9H, 2SiMe₃); ¹³C-NMR δ 138.85, 128.34-127.47 (2Ph), 99.19 (C-1), 80.56, 75.03, 74.60, 72.81, 72.00, 68.37, 62.84, 54.61 (OMe), 0.76 (2SiMe₃). Anal. Calcd. for C₂₇H₄₂O₆Si₂: C, 62.43; H, 8.15. Found: C, 62.29; H, 8.24.

Cyclohexyl 2,3-di-O-benzyl-4,6-bis-O-trimethylsilyl-β-D-glucopyranoside (**4g**): Yield 88%; mp 64.5-65.5°C; $[\alpha]_D$ +22.2° (c 1.0, CHCl₃); IR v_{max} (NaCl) 1250 cm⁻¹ (SiMe); ¹H-NMR δ 7.39-7.21 (m, 10H, 2Ph), 4.98, 4.73 (ABq, J 11.2 Hz, PhCH₂), 4.97, 4.62 (ABq, J 11.2 Hz, PhCH₂), 4.57-4.44 (m, 1H, H-1), 3.95-3.11 (m, 7H), 2.13-1.11 (m, 10H, 5CH₂), 0.12, 0.09 (each s, each 9H, 2SiMe₃); ¹³C-NMR δ 139.34, 138.90, 128.50-127.36 (2Ph), 102.28 (C-1), 85.11, 82.89, 77.69, 76.87, 75.52, 74.98, 71.13, 62.35 (C-6), 34.08, 32.29, 25.95, 24.32, 0.76 (2SiMe₃). Anal. Calcd. for C₃₂H₅₀O₆Si₂: C, 65.49; H, 8.59. Found: C, 65.35; H, 8.56.

Cyclohexyl 2,3-di-O-benzyl-4,6-bis-O-trimethylsilyl-β-D-mannopyranoside (6g): Yield 86%; mp 56.5-58.0°C; $[\alpha]_D$ -58.8° (c 1.1, CHCl₃); IR v_{max} (NaCl) 1240 cm⁻¹ (SiMe); ¹H-NMR δ 7.52-7.12 (m, 10H,

2Ph), 4.97, 4.73 (ABq, J 12.8 Hz, PhCH₂), 4.51, 4.37 (ABq, J 11.8 Hz, PhCH₂), 4.48 (s, 1H, H-1), 4.04-3.08 (m, 7H), 2.07-1.08 (m, 10H, 5CH₂), 0.14, 0.11 (each s, each 9H, 2SiMe₃); ¹³C-NMR δ 139.66, 138.63, 128.34-127.20 (2Ph), 99.68 (C-1), 82.67, 78.44, 76.28, 75.14, 74.00, 71.35, 68.58, 63.06, 33.70, 31.80, 26.00, 24.05, 23.95, 0.81 (2SiMe₃). Anal. Calcd. for C₃₂H₅₀O₆Si₂: C, 65.49; H, 8.59. Found: C, 65.05; H, 8.29.

Methyl 2,3-di-O-benzyl-6-O-(tert-butyldimethylsilyl)-4-O-trimethylsilyl- α -D-galactopyranoside (2f): To a solution of *tert*-butylchlorodimethylsilane (287 mg, 1.91 mmol) and imidazole (134 mg, 1.97 mmol) in dry DMF (1.75 ml) was added methyl 2,3-di-O-benzyl- α -D-galactopyranoside²⁶ (2a, 474 mg, 1.27 mmol). The starting material 2a disappeared after 1 h to give a mono-TBDMS derivative 2d exclusively and then to the mixture was added a solution of chlorotrimethylsilane (0.24 ml, 1.91 mmol) and imidazole (134 mg, 1.96 mmol) in dry DMF (1.75 ml). After 5 min, the mixture was concentrated and the residue was purified on a column of silica gel with hexane-ethyl acetate to give 2f (639 mg, 90 %) as a syrup; $[\alpha]_D$ +29.7° (c 1.8, CHCl₃); IR ν_{max} (NaCl) 1252 cm⁻¹ (SiMe); ¹H-NMR δ 7.44-7.24 (m, 10H, 2Ph), 4.85, 4.70 (ABq, J 12.0 Hz, PhCH₂), 4.81, 4.64 (ABq, J 12.4 Hz, PhCH₂), 4.70 (d, $J_{1,2}$ 3.8 Hz, H-1), 4.16-3.60 (m, 6H), 3.38 (s, 3H, OMe), 0.91 (s, 9H, CMe₃), 0.09 (s, 15H, 5SiMe). Anal. Calcd. for C₃₀H₄₈O₆Si₂: C, 64.24; H, 8.63. Found: C, 64.17; H, 8.66.

The mono-TBDMS derivative 2d was characterized as its 4-acetate 2e: ¹H-NMR δ 7.42-7.08 (m, 10H, 2Ph), 5.59 (d, 1H, J 2.6 Hz, H-4), 4.84, 4.65 (ABq, J 11.2 Hz, PhCH₂), 4.78, 4.56 (ABq, J 11.1 Hz, PhCH₂), 4.68 (d, $J_{1,2}$ 4.0 Hz, H-1), 4.00-3.58 (m, 5H), 3.39 (s, 3H, OMe), 2.10 (s, 3H, OAc), 0.88 (s, 9H, CMe₃), 0.04 (s, 6H, SiMe₂).

Methyl 2,3-di-O-benzyl-6-O- (*tert*-butyldimethylsilyl)-4-O-trimethylsilyl- β -D-galactopyranoside (5f): To a solution of *tert*-butylchlorodimethylsilane (0.73 g, 4.84 mmol) and imidazole (336 mg, 4.94 mmol) in dry DMF (5.0 ml) was added methyl 2,3-di-O-benzyl- β -D-galactopyranoside²⁷ (5a, 598 mg, 1.60 mmol). The starting material (5a) disappeared after 50 min to give a mono-TBDMS derivative 5d and then to this mixture was added a solution of chlorotrimethylsilane (0.30 ml, 2.4 mmol) and imidazole (168 mg, 2.47 mmol) in dry DMF (2.5 ml). After 10 min, the mixture was concentrated and the residue was purified on a column of silica gel with hexane-ethyl acetate to give 5f (885 mg, 95%) as a syrup; [α]_D +8.1° (c 0.8, CHCl₃); IR v_{max} (NaCl) 1252 cm⁻¹ (SiMe); ¹H-NMR δ 7.41-7.21 (m, 10H, 2Ph), 4.75, 4.69 (ABq, J 11.5 Hz, PhCH₂), 4.71, 4.23 (s, 2H, PhCH₂), 4.23 (d, J_{1,2} 7.6 Hz, H-1), 4.11-3.23 (m, 6H), 3.55 (s, 3H, OMe), 0.90 (s, 9H, CMe₃), 0.09 (s, 15H, SiMe₂, SiMe₃). Anal. Calcd. for C₃₀H₄₈O₆Si₂: C, 64.24; H, 8.63. Found: C, 63.90; H, 8.46.

The mono-TBDMS derivative 5d was characterized as its 4-acetate 5e: ¹H-NMR δ 7.40-7.10 (m, 10H, 2Ph), 5.54 (br d, 1H, H-4), 4.86, 4.70 (ABq, J 10.5 Hz, PhCH₂), 4.79, 4.51 (ABq, J 11.5 Hz, PhCH₂), 4.90-3.60 (m, 6H), 3.58 (s, 3H, OMe), 2.13 (s, 3H, OAc), 0.91 (s, 9H, CMe₃), 0.06 (s, 6H, SiMe₂).

General method of pyruvate acetalization. Thoroughly dried bis-silylated diol (1.0 mmol) and ethyl or benzyl pyruvate (2.0 mmol) were dissolved in dichloromethane or diethyl ether (1 ml). After the addition of TMSOTF (0.4 mmol) in the solvent (0.1 ml) at -20°C this solution was kept at the specified temperature for several days as shown in Tables 1 and 3 to complete the reaction. Pyridine (1 ml) was added to this solution. The mixture was poured into cold aqueous sodium hydrogencarbonate (15 ml) and extracted with chloroform (15 ml x 3). The extract was washed with water, dried over anhydrous sodium sulfate, and evaporated. The residue was subjected to column chromatography on silica gel with hexane-ethyl acetate to give pyruvate acetal. Reaction conditions and yields are given in the tables.

Methyl 2,3-di-O-benzyl-4,6-O-{(R)- and (S)-(1-ethoxycarbonyl)ethylidene}- α -D-glucopyranoside (13R and 13S). On chromatographic separation of two isomers hexane-ethyl acetate mixtures of the following ratios were used as eluant: 13S (7:1); 13R (5:1 to 4:1).

13R: mp 99.0-100.0 °C; $[\alpha]_D$ -9.6° (c 1.0, CHCl₃); IR v_{max} (NaCl) 1720 cm⁻¹ (ester); ¹H-NMR δ 7.49-7.25 (m, 10H, 2Ph), 4.90, 4.75 (ABq, J 11.0 Hz, PhCH₂), 4.84, 4.75 (ABq, J 12.0 Hz, PhCH₂), 4.53 (d, J_{1,2} 4.0 Hz, H-1), 4.23 (q, 2H, J 7.0 Hz, CH₂Me), 4.11-3.50 (m, 5H), 3.50 (dd, J_{2,3} 9.0 Hz, H-2), 3.35 (s, 3H, OMe), 1.67 (s, 3H, CMe), 1.31 (t, 3H, J 7.0 Hz, CH₂Me); ¹³C-NMR δ 168.43 (C=O), 138.25, 128.45-127.58 (2Ph), 99.41 (C-1), 97.68 (acetal), 78.93, 78.50, 74.81, 62.73 (C-2,3,4,5), 75.14, 73.84 (2PhCH₂), 63.28, 61.87 (C-6, CH₂Me), 55.37 (OMe), 17.88 (CMe), 14.09 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 66.02; H, 6.71.

13S: $[α]_D$ +44.8° (c 1.0, CHCl₃); IR ν_{max} (NaCl) 1736 cm⁻¹ (ester); ¹H-NMR δ 7.60-7.17 (m, 10H, 2Ph), 4.96, 4.77 (ABq, J 11.0 Hz, PhCH₂), 4.81, 4.61 (ABq, J 12.0 Hz, PhCH₂), 4.50 (d, $J_{1,2}$ 3.5 Hz, H-1), 4.25 (q, 2H, J 7.5 Hz, CH₂Me), 4.05-3.28 (m, 6H), 3.34 (s, 3H, OMe), 1.55 (s, 3H, CMe), 1.30 (t, 3H, CH₂Me); ¹³C-NMR δ 169.73 (C=O), 139.07, 138.31, 128.34-127.36 (2Ph), 99.25 (C-1), 98.98 (acetal), 78.82, 78.55, 78.28 (C-2,3,4,5), 74.60, 73.84 (2PhCH₂), 65.50, 61.70 (C-6, CH₂Me), 55.37 (OMe), 25.46 (CMe), 14.19 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 66.04; H, 6.90.

Methyl 2,3-di-O-benzyl-4,6-O- $\{(R)$ - and (S)- $(1-ethoxycarbonyl)ethylidene}-<math>\alpha$ -D-galactopyranoside (14R and 14S). On chromatographic separation of two isomers hexane-ethyl acetate mixtures of the following ratios were used as eluant: 14R (3:1); 14S (2:1).

14R: $[α]_D$ +48.4° (c 1.0, CHCl₃); IR v_{max} (NaCl) 1720 cm⁻¹ (ester); ¹H-NMR δ 7.50-7.10 (m, 10H, 2Ph), 4.91, 4.69 (ABq, J 12.0 Hz, PhCH₂), 4.87, 4.66 (ABq, J 12.0 Hz, PhCH₂), 4.71 (d, J_{1,2} 3.8 Hz, H-1), 4.30 (q, 2H, J 7.0 Hz, CH₂Me), 4.23 (m, 1H, H-6a), 4.12-3.79 (m, 4H), 3.48 (m, H-5), 3.35 (s, 3H, OMe), 1.64 (s, 3H, CMe), 1.36 (t, 3H, CH₂Me); ¹³C-NMR δ 170.00 (C=0), 138.69, 138.42, 128.34-127.58 (2Ph), 99.52 (C-1), 98.76 (acetal), 75.19, 75.03, 69.61, 61.49 (C-2,3,4,5), 73.95, 71.13 (2PhCH₂), 65.44, 61.70 (C-6, CH₂Me), 55.53 (OMe), 26.00 (CMe), 14.30 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 66.40; H, 7.03.

14S: $[α]_D$ +70.4° (c 1.0, CHCl₃); IR v_{max} (NaCl) 1720 cm⁻¹ (ester); ¹H-NMR δ 7.40-7.00 (m, 10H, 2Ph), 4.88, 4.65 (ABq, J 12.0 Hz, PhCH₂), 4.82, 4.64 (ABq, J 12.5 Hz, PhCH₂), 4.60 (d, $J_{1,2}$ 3.0 Hz, H-1), 4.25-3.76 (m, 7H), 3.63 (m, H-5), 3.31 (s, 3H, OMe), 1.56 (s, 3H, CMe), 1.21 (t, 3H, J 7.0 Hz, CH₂Me); ¹³C-NMR δ 168.97 (C=O), 138.69, 128.34-127.64 (2Ph), 99.25 (C-1), 97.19 (acetal), 75.74, 75.25, 67.66, 62.84 (C-2,3,4,5), 73.84, 72.38 (2PhCH₂), 63.98, 61.54 (C-6,CH₂Me), 55.53 (OMe), 20.37 (CMe), 14.03 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 66.02; H, 6.95. Methyl 2,3-di-O-benzyl-4,6-O- $\{(R)$ - and (S)-(1-ethoxycarbonyl)ethylidene $\}$ - α -D-mannopyranoside (15R and 15S). On chromatographic separation of two isomers hexane-ethyl acetate mixtures of the following ratios were used as eluant: 15S (8:1 to 7:1); 15R (6:1 to 5:1).

15 R: $[\alpha]_D$ +44.7° (c 1.1, CHCl₃); IR v_{max} (NaCl) 1730 cm⁻¹ (ester); ¹H-NMR δ 7.48-7.17 (m, 10H, 2Ph), 4.88-4.58 (m, 5H, H-1, 2PhCH₂), 4.42-3.52 (m, 8H), 3.26 (s, 3H, OMe), 1.72 (s, 3H, CMe), 1.32 (t, 3H, J 7.2 Hz, CH₂Me); ¹³C-NMR δ 168.59 (C=O), 138.96, 138.20, 128.39-127.42 (2Ph), 100.66 (C-1), 98.00 (acetal), 78.50, 76.28, 72.05, 64.36 (C-2,3,4,5), 73.68, 73.08 (2PhCH₂), 63.06, 61.81 (C-6, CH₂Me), 54.77 (OMe), 17.88 (CMe), 14.14 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 66.11; H, 6.92.

15 S: $[\alpha]_{D}$ +70.6° (c 1.0, CHCl₃); IR v_{max} (NaCl) 1735 cm⁻¹ (ester); ¹H-NMR & 7.44-7.20 (m, 10H, 2Ph), 4.96-4.66 (m, 5H, H-1, 2PhCH₂), 4.44-3.50 (m, 8H), 3.26 (s, 3H, OMe), 1.52 (s, 3H, CMe), 1.30 (t, 3H, J 7.0 Hz, CH₂Me); ¹³C-NMR & 169.95 (C=O), 139.23, 138.42, 128.18-127.31 (2Ph), 100.82 (C-1), 99.30 (acetal), 76.28, 75.90, 75.68, 63.60 (C-2,3,4,5), 73.73, 72.97 (2PhCH₂), 65.39, 61.70 (C-6, CH₂Me), 54.82 (OMe), 25.62 (CMe), 14.19 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 65.86; H, 6.98.

Cyclohexyl 2,3-di-O-benzyl-4,6-O- $\{(R)$ - and (S)- $(1-ethoxycarbonyl)ethylidene}-\beta-D-gluco$ pyranoside (17R and 17S). On chromatographic separation of two isomers hexane-ethyl acetate mixturesof the following ratios were used as eluant: 17S (10:1); 17R (9:1 to 8:1).

17R: $[\alpha]_{D}$ -16.8° (c 1.1, CHCl₃); IR v_{max} (NaCl) 1735 cm⁻¹ (ester); ¹H-NMR δ 7.44-7.20 (m, 10H, 2Ph), 4.88, 4.71 (ABq, J 14.0 Hz, PhCH₂), 4.86, 4.71 (ABq, J 15.0 Hz, PhCH₂), 4.53 (d, $J_{1,2}$ 7.6 Hz, H-1), 4.25 (q, 2H, J 7.6 Hz, CH₂Me), 4.11 (dd, $J_{5,6a}$ 6.0 Hz, J_{gem} 11.2 Hz, H-6a), 3.98-3.24 (m, 6H), 2.08-1.08 (m, 10H, 5CH₂), 1.68 (s, 3H, CMe), 1.32 (t, 3H, CH₂Me); ¹³C-NMR δ 168.43 (C=O), 138.69, 138.52, 128.18-127.58 (2Ph), 102.34 (d, J_{C-H} 158.7 Hz, C-1), 97.51 (acetal), 82.02, 80.83, 78.06, 74.16, 66.36, 75.30, 74.81 (2PhCH₂), 63.00 (CH₂Me), 61.83 (C-6), 33.70, 31.90, 25.57, 24.05, 23.89, 17.82 (CMe), 14.08 (CH₂Me). Anal. Calcd. for C₃₁H₄₀O₈: C, 68.87; H, 7.46. Found: C, 68.47; H, 7.57.

17S: $[\alpha]_{D}$ +25.0° (c 1.1, CHCl₃); IR v_{max} (NaCl) 1735 cm⁻¹ (ester); ¹H-NMR δ 7.47-7.14 (m, 10H, 2Ph), 4.93, 4.72 (ABq, J 11.8 Hz, PhCH₂), 4.85, 4.66 (ABq, J 10 Hz, PhCH₂), 4.52 (d, J_{1,2} 8.0 Hz, H-1), 4.24 (q, 2H,J 7.2 Hz, CH₂Me), 4.04 (dd, J_{5,6a} 5.4 Hz, J_{gem} 10.6 Hz, H-6a), 3.88-3.16 (m, 7H), 2.04-1.08 (m, 10H, 5CH₂), 1.55 (s, 3H, CMe), 1.30 (t, 3H, CH₂Me); ¹³C-NMR δ 169.67 (C=O), 138.96, 138.58, 128.23-127.36 (2Ph), 102.17 (d, J_{C-H} 158.7, C-1), 98.92 (acetal), 81.91, 80.88, 77.90, 65.50, 75.41, 74.22 (2PhCH₂), 65.23 (CH₂Me), 61.76 (C-6), 33.70, 31.91, 25.57, 24.00, 23.89, 25.46 (CMe), 14.25 (CH₂Me). Anal. Calcd. for C₃₁H₄₀O₈: C, 68.87; H, 7.46. Found: C, 68.72; H, 7.56.

Methyl 2,3-di-O-benzyl-4,6-O- $\{(R) \text{ and } (S)-(1-\text{ethoxycarbonyl})\text{ethylidene}\}-\beta-D-galactopyra$ noside (18R and 18S). The two isomer were separated by elution with 3:1 mixture of hexane and ethylacctate.

18R: $[\alpha]_{D}$ +12.8° (c 1.0, CHCl₃); IR ν_{max} (NaCl) 1735 cm⁻¹ (ester); ¹H-NMR δ 7.48-7.20 (m, 10H, 2Ph), 4.88, 4.62 (ABq, J 12.2 Hz, PhCH₂), 4.84 (s, 2H, PhCH₂), 4.31 (q, 2H, J 7.2 Hz, CH₂Me), 4.29-3.18 (m,

6H), 3.79 (dd, $J_{1,2}$ 8.0 Hz, $J_{2,3}$ 10.0 Hz, H-2), 3.56 (s, 3H, OMe), 1.65 (s, 3H, CMe), 1.34 (t, 3H, CH₂Me); ¹³C-NMR δ 170.11 (C=O), 138.90, 138.09, 128.29-127.47 (2Ph), 104.61 (C-1), 98.76 (acetal), 78.50, 78.17, 68.58, 65.61 (C-2,3,4,5), 75.30, 70.81 (2PhCH₂), 65.33, 61.65 (C-6, CH₂Me), 57.21 (OMe), 25.90 (CMe), 14.30 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 65.99; H, 6.94.

18S: $[α]_D$ +12.2° (c 1.0, CHCl₃); IR v_{max} 1715 cm⁻¹ (ester); ¹H-NMR δ 7.48-7.20 (m, 10H, 2Ph), 4.89, 4.78 (ABq, J 10.2 Hz, PhCH₂), 4.76 (s, 2H, PhCH₂), 4.22 (q, 2H, J 7.2 Hz, CH₂Me), 4.24-3.31 (m, 6H,), 3.85 (dd, $J_{1,2}$ 8.0 Hz, $J_{2,3}$ 10.0 Hz, H-2), 3.50 (s, 3H, OMe), 1.58 (s, 3H, CMe), 1.33 (t, 3H, CH₂Me); ¹³C-NMR δ 169.51(C=O), 138.90, 138.40 128.34-127.47 (2Ph), 104.18 (C-1), 96.97 (acetal), 79.09, 78.49, 67.23, 66.63 (C-2,3,4,5), 75.25, 72.00 (2PhCH₂), 64.85, 61.54 (C-6, CH₂Me), 56.77 (OMe), 23.35 (CMe), 13.98 (CH₂Me). Anal. Calcd. for C₂₆H₃₂O₈: C, 66.08; H, 6.83. Found: C, 65.26; H, 7.04.

Pyruvate acetalization of β -D-mannopyranoside 6g. The acetalization of 6g in CH₂Cl₂ (Table 1, entry 11) gave a mixture of anomerized 4,6-acetals 16R and 16S, and 5,6-acetal 20, which were separated by elution with hexane-ethyl acetate (8:1 to 5:1), and characterized only by ¹H-NMR and ¹³C-NMR.

Cyclohexyl 2,3-di-O-benzyl-4,6-O-{(R)-(1-ethoxycarbonyl)ethylidene')- α -D-mannopyranoside (16R): ¹H-NMR δ 7.44-7.20 (m, 10H, 2Ph), 4.92-4.51 (m, 5H, 2PhCH₂, H-1), 4.28 (q, 2H, J 7.0 Hz, CH₂Me), 4.36-3.36 (m, 7H), 2.0-1.0 (m, 10H, 5CH₂), 1.72 (s, 3H, CMe), 1.26 (t, 3H, CH₂Me); ¹³C-NMR δ 168.59 (C=O), 138.07, 128.18-127.42 (2Ph), 97.89 (acetal), 97.46 (d, J_{C-H} 168.46, C-1), 77.36, 76.55, 75.14, 73.68, 73.24, 72.32, 63.06, 61.76, 64.52, 33.26, 31.15, 25.57, 23.95, 23.73,17.82 (CMe), 14.08 (CH₂Me). Its (S)-isomer 16S: ¹H-NMR δ 7.47-7.17 (m, 10H, 2Ph), 4.97, 4.75 (ABq, J 12.0 Hz, PhCH₂), 4.82, 4.59 (ABq, J 12.0 Hz, PhCH₂), 4.59 (s, 1H, H-1), 4.45-3.68 (m, 9H), 2.03-1.05 (m, 10H, 5CH₂), 1.57 (s, 3H, CMe), 1.37 (t, 3H, CH₂Me); ¹³C-NMR δ 169.95 (C=O), 139.34, 128.18-127.20 (2Ph), 99.25 (acetal), 97.62 (d, J_{C-H} 168.5 Hz, C-1), 63.76 (QCH), 25.57 (CMe), 14.13 (CH₂Me).

Cyclohexyl 2,3-di-O-benzyl-5,6-O-{(1-ethoxycarbonyl)ethylidene}-D-mannopyranoside (20): ¹H-NMR δ 7.48-7.20 (m, 10H, 2Ph), 5.22 (d, $J_{1,2}$ 3.6 Hz, H-1), 4.75, 4.66 (ABq, J 11.8 Hz, PhCH₂), 4.63, 4.59 (ABq, J 12.0 Hz, PhCH₂), 4.47 (dt, $J_{5,6a}$ 6.0 Hz, $J_{5,6b}$ 7.8 Hz, $J_{5,4}$ 7.5 Hz, H-5), 4.24 (dd, $J_{4,3}$ 4.5 Hz, H-4), 4.22 (dd, J_{gem} 8.8 Hz, H-6a), 4.20 (dd, H-6b), 3.90 (dd, H-2), 3.55-3.48 (m, 1H, OCH), 1.58 (s, 1H, CMe), 1.27 (t, 3H, CH₂Me), 1.92-1.12 (m, 10H, 5CH₂); ¹³C-NMR δ 170.32 (C=O), 138.42, 128.34-127.58 (2Ph), 105.59 (acetal), 104.23 (C-1), 83.81, 79.15, 77.96, 76.28, 74.71, 73.68, 72.49, 68.91, 61.27, 33.64, 31.96, 25.68, 24.00, 22.70 (CMe), 14.09 (CH₂Me).

The acetalization of 6 g in Et₂O (Table 3, entry 9) gave the desired 4,6-acetals 19 R in low yields together with 20, which were separated by elution with hexane-ethyl acetate (10:1 to 8:1) and characterized only by ¹H-NMR and ¹³C-NMR.

Cyclohexyl 2,3-di-O-benzyl-4,6-O-{(R)-(1-ethoxycarbonyl)ethylidene}- β -D-mannopyranoside (19R):¹H-NMR δ 7.52-7.18 (m, 10H, 2Ph), 4.97, 4.84 (ABq, J 12.0 Hz, PhCH₂), 4.69, 4.54 (ABq, J 11.8 Hz, PhCH₂), 4.48 (s, 1H, H-1), 4.25 (q, 2H, J 7.0 Hz, CH₂Me), 4.41-3.18 (m, 7H), 2.00-1.04 (m, 10H, 5CH₂), 1.70 (s, 3H, CMe), 1.32 (t, 3H, CH₂Me); ¹³C-NMR δ 168.50 (C=O), 138.63, 128.66-127.42 (2Ph), 100.06 (d, J_{C-H} 152.6 Hz, C-1), 97.95 (acetal), 78.11, 76.71, 76.55, 74.59, 72.32, 71.62, 67.93, 62.84, 61.76, 33.37, 31.42, 25.68, 23.73, 23.57, 17.93 (CMe), 14.14 (CH₂Me).

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- 13. Coupling of the disilyl ether 3 c with benzyl pyruvate at 3 °C in dichloromethane or ether gave, albeit low yield (22–25%), the corresponding (S)-acetal exclusively. Methyl 2,3-di-O-benzyl-4,6-O-{(S)-(1-benzyloxycarbonyl) ethylidene}- α -D-mannopyranoside: [α]_D +70.3° (c 1.0, CHCl₃); IR v_{max} (NaCl) 1735 cm⁻¹ (ester); ¹H-NMR δ 7.50-6.90 (m, 5H, 3Ph), 5.28, 5.11 (ABq, J 12.5 Hz, PhCH₂), 4.88, 4.64 (ABq, J 12.0 Hz, PhCH₂), 4.68-4.46 (m, 3H, H-1, PhCH₂), 4.26-3.51 (m, 6H), 3.25 (s, 3H, OMe), 1.57 (s, 3H, CMe); ¹³C-NMR δ 169.84 (C=O), 139.01, 138.31, 135.27, 128.83-126.82 (3Ph), 100.66 (C-1), 99.30 (acetal), 76.44, 76.33, 75.90, 63.44 (C-2,3,4,5), 73.57, 73.03, 67.28, 65.23 (3PhCH₂, C-6), 54.72 (OMe), 25.57 (CMe). Anal. Calcd. for C₃₁H₃₄O₈: C, 69.64; H, 6.54. Found: C, 69.75; H, 6.42.
- 14. The stereoselectivities in the acetalization of both gluco isomers (1 c and 4 c) in dichloromethane cannot be explained solely by thermodynamic factor. The acetalization of the α-manno isomer 3 c in the presence of TMSOTF (40 mol%) and TfOH (10 mol%) in dichloromethane at -5 °C for 1 day afforded the (R)-acetal 15R and the (S)-acetal 15S in yields of 18% and 45%, respectively, together with a small amount (6%) of the 5,6-acetal 20. Thus, the acetalization reaction at the lower temperature may proceed under kinetic control.
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- The acetalization of 1g in CH₂Cl₂ or in ether containing 40 mol% TMSOTf gave, even at lower temperatures (-50 -30 °C), mainly the intermolecular 6,6'-acetal I in 15 39% yield. The formation of I suggests that hydrolysis of 4-O-TMS ether competes with the 4,6-acetal ring formation. Ethyl pyruvate bis-(methyl 2,3-di-O-benzyl-α-D-glucopyranoside-6-yl)acetal (I). [α]_D +14.9° (c 1.0, CHCl₃); IR v_{max} (NaCl) 3600-3100 (OH), 1715 cm⁻¹ (ester); ¹H-NMR δ 7.44-7.20 (m, 20H, 4Ph), 4.97, 4.80 (ABq, 4H, J 12.0 Hz, 2PhCH₂), 4.76, 4.60 (ABq, 4H, J 12.0 Hz, 2PhCH₂), 4.76, 4.60 (ABq, 4H, J 12.0 Hz, 2PhCH₂), 4.62 (d, 2H, J_{1,2} 3 Hz, H-1,1'), 4.20 (q, 2H, J 7.0 Hz, CH₂Me), 3.96-3.40 (m, 12H), 3.37 (s, 6H, 2OMe), 3.32-2.88 (br, 2H, 2OH), 1.55 (s,

3H, CMe), 1.28 (t, 3H, CH₂Me); ¹³C-NMR δ 169.57 (C=O), 99.84 (acetal), 98.11 (C-1), 21.62 (CMe), 14.03 (CH₂Me). Anal. Calcd. for C₄₇H₅₈O₁₄: C, 66.65; H, 6.90. Found: C, 66.80; H, 6.93.The acetal I was further characterized as its 4, 4'-di-O-acetyl derivative II: ¹H-NMR δ 7.35-7.26 (m, 10H, 2Ph), 4.87, 4.64 (ABq, J 12.0 Hz, PhCH₂), 4.77, 4.63 (ABq, J 12.0 Hz, PhCH₂), 4.77, 4.63 (ABq, J 12.0 Hz, PhCH₂), 4.85, 4.83 (each t, J_{3,4} = J_{4,5} 10.0 Hz, H-4, H-4'), 4.59 (d, J_{1,2} 3.4 Hz, H-1), 4.58 (d, J_{1,2} 3.9 Hz, H-1'), 4.17 (q, J 7.0 Hz, CH₂Me), 3.87, 3.88 (each t, J_{2,3} = J_{3,4} 9.4 Hz, H-3, H-3'), 3.56-3.39 (m, 8H), 3.38, 3.37 (each s, each 3H, 2OMe), 1.90, 1.86 (each s, each 3H, OAc), 1.45 (s, 3H, CMe), 1.24 (t, 3H, CH₂Me).



- 18. An attempt to prevent the ring contraction and anomerization employing the more bulky Lewis acid, TBDMSOTf instead of TMSOTf was unsuccessful. While the acetalization of 3g in the presence of TBDMSOTf (40 mol%) in Et₂O gave only the 4,6-acetal 15, the yield was rather low [14% (R)-isomer and 11% (S)-isomer]. Further, the acetalization of 6g under the same conditions gave an anomeric mixture (15% yield) of the 4,6-acetal.
- 19. In the presence of additional TfOH (2 mol%) the reaction proceeded at -20 °C to afford 15R and 15S in yields of 31% and 28%, respectively. The induction time is deduced to be necessary for the formation of potential actual catalyst TfOH by decomposition of TMSOTF.
- 20. The data given for 25b in the communication¹⁰ including yields and 1H- and 13C-NMR chemical shifts were mistaken for those of 18S and 18R.
- 21. The structure of 27 was deduced by ¹³C-NMR chemical shift of the acetal carbon (98.02 ppm) indicating 6-membered ring and by ¹H-NMR chemical shift of methylene in ethoxyl group, which resonates at the higher field (3.51 ppm) than that in ethyl ester (ca. 4.2 ppm). Further, the lactone structure was supported by the ¹H-detected heteronuclear multiple-bond correlation (HMBC) measurement, which confirmed the correlation between H-3 and carbonyl carbon. A possible mechanism for the formation of 27 may be shown as follows.



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